

Heavy Metal Contaminants Removal from Wastewater Using the Potential Filamentous Fungi Biomass: A Review

Shafiquzzaman Siddiquee^{1*}, Kobun Rovina¹, Sujjat Al Azad², Laila Naher³, Saallah Suryani¹ and Pasicha Chaikaew⁴

¹Biotechnology Research Institute, Universiti Malaysia Sabah, Jln UMS, Sabah, Malaysia

²Borneo Marine Research Institute, Universiti Malaysia Sabah, Jln UMS, Sabah, Malaysia

³Faculty of Agro Based Industry, Universiti Kelantan Malaysia, Kelantan

⁴Department of Environmental Science, Faculty of Science, Chulalongkorn University, Bangkok, Thailand

Abstract

Heavy metal pollution of wastewater currently becomes a key environmental problem throughout the whole world. Conventional methods for the removal of heavy metals from aqueous solutions are not economically and environmental friendly because it has produced massive quantity of toxic chemical compounds. Recently, the removal of heavy metals from wastewater are extensively used various conventional methods such as chemical precipitation, coagulation-flocculation, flotation, ion exchange and membrane filtration. Biological treatments, especially filamentous fungi have gained an increasing attention for heavy metal removal and recovery due to their upright performances, low cost and huge quantities. The filamentous fungi have a great potential to produce large amount of biomasses which are widely used for metal adsorption capacities of Pb, Zn, Cd, Cu, Cr, As and Ni. Production of biomass has offered great potential for adopting metal-recovery system. The main aim of this review paper is to discuss the available information of heavy metals removal for the utilization of filamentous fungi biomass and scrutinize the practical of exploiting them for heavy metal remediation.

Keywords: Heavy metals; Filamentous fungi; Wastewater; Bio sorption; Biomass

Introduction

The soil and water contaminations are frequently occurred by toxic heavy metals and organic pollutants as a consequence of human activities become a key concern in environmental and health problem. Several toxic metals (Cd, Cu, Hg, Pb, Mn, As, Ni, Zn, etc.) from industrial wastewater and other human activities are directly or indirectly released into the environment. Unlike organic contaminants, these pollutants from heavy metals are not biodegradable and able to travel up the food chain via bioaccumulation. According to Lopez and Vazquez [1], some metals including Cu, Fe, Mn and Zn are micro-nutrients for most of the organisms; however, not all living organism. It can play a vital role in metalloenzymes. Cations usually increase membrane stability and play specific roles in nucleic acid structures, functions and metabolisms [2]. When the concentrations of beneficial metals (mercury, lead, cadmium) in the environment are very high, they become more toxic [2,3].

Volesky [4] and Domenech [5] shortlisted some available conventional methods for removing the dissolved heavy metals including chemical precipitation and sludge separation, chemical oxidation or reduction, ion exchange, reverse osmosis, filtration, adsorption using activated charcoal, electrochemical treatment and evaporative recovery. These physio-chemical techniques are costly and may not always be practicable, and also their metal-binding properties are non-specific [6]. The removal of heavy metals is more taken into consideration when using microbial biomass [4].

Naturally fungi have a large variety of extracellular proteins, organic acids and other metabolites. Fungi can adapt in any ecosystems and any environmental conditions [7]. Most of the soil fungi, especially, the filamentous fungi of *Trichoderma* are a great interest in agriculture. *Trichoderma* species is commonly found in root, soil, foliar environments and all environmental conditions. It can grow faster, high strong spore producers, cell wall degrading

enzymes, biocontrol agent and eco-friendly in nature [8]. It has high resistant towards various toxins and xenobiotic compounds, including antibiotics, fungicides and heavy metals [9]. Benítez et al. [10] have reported that some potential *Trichoderma* strains are possessed good antagonistic abilities against some plant pathogenic fungi. Kredics et al. [11,12] have suggested that only mercury (Hg) do not cause any inhibition of extracellular enzyme production in the mycoparasite. For that reason, *T. harzianum* is more effective under heavy metal stress condition towards mycoparasitisms.

Heavy Metals

Heavy metals are defined on the basis of three different criteria which including density, atomic number or their chemical properties. Radojevic et al. [13] have mentioned that heavy metals can be defined by density of such metals (5 g/cm³) which is denser than water by five times. Yu [14] categorized metals as any elements with an atomic number of higher than 20 excluding alkaline metals, alkaline earth, lanthanides and actinides. In sewage water, concentration of Zn is found highest value compared to other heavy metals as shown in Table 1 [15].

Pollution and disease caused by heavy metals

The existing heavy metals in the environment and industrial

***Corresponding authors:** Biotechnology Research Institute, Universiti Malaysia Sabah, Jln UMS, Sabah, Malaysia, Tel: 60 88-320 000; E-mail: shafiqpab@ums.edu.my

Received October 05, 2015; **Accepted** October 30, 2015; **Published** November 06, 2015

Citation: Siddiquee S, Rovina K, Azad SA, Naher L, Suryani S, et al. (2015) Heavy Metal Contaminants Removal from Wastewater Using the Potential Filamentous Fungi Biomass: A Review. J Microb Biochem Technol 7: 384-393. doi:10.4172/1948-5948.1000243

Copyright: © 2015 Siddiquee S, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Type of metals	Concentration (mgL ⁻¹)
Cd	10
Ni	60
Cr	240
Pb	450
Cu	700
Zn	2600

Table 1: Metals concentration (mgL⁻¹) in sewage [15].

wastewater increasingly pollute ecosystems and threaten human health in-the developing countries. Different concentrations of heavy metal elements commonly occur in all ecosystems. Several compounds have diverse properties such as Zn, Cu, Ni, Fe and Mn are essential trace elements for living organisms [16]. The high level accumulation of these metals or ingested in greater amounts than the required concentration can produce serious problem towards living things including human being. Alleviating of heavy metal concentration in water is crucial to the quality of aquatic living organisms. In addition, heavy metals can cause severe toxic effects to expose plants, animals and human when presence in the excessive concentrations [16]. The wastes containing metals are directly or indirectly being discharged into the environment producing serious environmental pollution and possess a major threat toward human, soil and sediments health [17,18].

Arora et al. [19] have reported that heavy metals have the ability to accumulate in different parts of human body. Heavy metal concentrations have the characteristic of having long biological half-lives as well as resistant to degradable process and exhibit their chemical toxicity in soluble water. For this reason, the heavy metals are considered as a threat to human and other organisms even though when present in low concentration. When the concentration of heavy metals is entered the human body through absorption, these metals ion can bind various biomolecules such as proteins, nucleic acid and interfere with their functions [14].

Contaminations of heavy metals in environments

Rapid developments of industrializations and urbanizations have led to the direct impact of the environment. The resultant degradation and contamination of ecosystem become a major threat toward all living organisms worldwide, in particular, human beings. Globally, open water and aquatic ecosystems are contaminated with several heavy metals through various human activities that indirectly or directly lead to these pollutions [20]. The most serious water pollutions have occurred with some water bodies such as rivers, lakes, oceans and groundwater. In additions, high amount of materials can change the water properties and polluted the water, thus resulting in unfit for intended uses. The water pollution can be classified into two distinct types that are point sources and nonpoint sources [20].

The point sources of pollutions are single identifiable localized sources of water pollution. It can occur when harmful substances are emitted directly into the water bodies. Examples of water pollution are the Exxon Valdez oil spills in 1989 that ran aground in the Price William Sound causing expelling of 11 million gallons of crude oil into the Alaska environment [20]. The nonpoint source pollution is affecting the water body from the diffusion sources such as polluted runoff from agricultural areas draining into the river and oceans waters. Besides, the nonpoint sources pollutions are derived and come from many different sources which made it difficult to find the specific solution to stop the pollution.

Types and toxicity effects of heavy metals

Zinc: Zinc (Zn) covers about 0.004% of the earth crust with atomic number of 30 and atomic relative number of 65.39. Zn has the average content approximately 80 ppm in lithosphere and between 10–300 ppm in soil [21]. Zinc is a one of major metal that can be found in effluents discharged from industries [22,23]. Most of the industries are released zinc as their waste material which includes electroplating, manufacture of batteries, galvanization and metallurgical industries [23]. Zn can play an important role on plant growth. The growth slows down when Zn is deficient in the environment. Functional group of Zn can increase the stabilization of plant by altering its structure molecule and its membrane as well as act as the defensive mechanism against disease for the plant. Basically, zinc in metallic form does not cause any harm towards the environments and it has limited bioavailability. However, the presence of other chemicals such as acids and oxygen can react with zinc to form a potentially toxic compound which can cause severe damage to biological systems [24].

Copper: Copper is recognized another metal and contributed the heavy metals pollution or contamination. Copper (Cu) mostly found deep inside the earth. It is rare element which is found in nature in the un-combined state as well as in ores such as chalcopyrite [25]. According to Davies and Bennett [26], copper is classed as the transition element in periodic table and included among 25 elements that are found inside earth crusts that have the atomic number of 29 and atomic weight of 63.55. The pollution of Cu in environment may have caused by animal manure. After that, Cu is used as supplement for inhibition of parasites as well as the industrial factory such as paint factory and the disposed by electronic and electrical factory [27]. The anthropogenic sources of copper element is mostly found in pollution sites from landfills, mine sites, combustion of fossil fuels and domestic waste waters. It can be released by particles from volcanoes, dust and forest fire into atmosphere or dissolved compound in water.

The contamination of copper metals can be hazardous to human health. Its can affect human either by its toxicity or by chronic diseases. The contamination of copper can cause severe damage of kidney and liver and even death when consume in high concentrations. The accumulation of Cu in the organs and the toxicity inside human bodies are increased with the decreasing of zinc element and sulphate ion [28] and also threaten the fish population. When the presence of heavy metal is highly found in fish's body, it can adversely alter the functional organ system. [29]. Even though copper play important activities in several enzymes for production of haemoglobin, it has detrimental effects towards some facts of the organisms. In living things, Cu is essential for living organisms as it acts as an antioxidant, participating in the electron transport chain as well as in collagen and elastin. This micronutrient is only required in certain amount which mostly accumulated inside human's tissues [30].

Nickel: Apart from Copper, nickel is known as major environment pollutant. It has possessed the potential of clastogenic, toxic and carcinogenic effect. The different solubility of nickel compound has different carcinogenic potentials. Dunnick et al. [31] have stated that insoluble Ni₃S₂ or NiO is strong carcinogens while the soluble nickel salts are presented as weak carcinogens. Nickel is commonly an airborne contaminant in the form of nickel carbonyl Ni(CO)₄, an intermediate product of refining nickel activity which enter the body through respiratory system. It has a high toxic volatile liquid that can cause death when inhaled and lead to several health problems such as pulmonary edema, pneumonia and respiratory failure [14]. Crosby [32] reported that some individuals have higher sensitivity towards nickel

compounds even having minimal contact with nickel plate or coins could result in irritation on their exposed part of body. The sources and the effects of heavy metals concentrations towards the environment are shown in Table 2 [22].

Heavy metals pollution cases: Accumulation of different types of heavy metals (e.g., Pb, Cd, Cu, Ni, Zn, and Mn) in the sea water not only contaminated the water but also occurred in soil. It is affecting the sources of drinking and building up the dangerous concentration of heavy metals in grains and vegetables. There are several cases involved with the heavy metals contaminations which occurred in 1963 in Minamata Bay, Japan. Its tragedy has related with local people who consumed shellfish that contained high amount of mercury concentration near the Minamata Bay. The exposure of these diseases occurred due to the chemical substances that released and discharged without controller by the chemical factory which operate near to the bay [33]. High amount of mercury concentration is discharged into the sea as wastewater and affect the marine foods chains such as shellfish and other seafood which can build high concentration of mercury than become poisonous to the locals who are consumed them [33].

Concentrations of heavy metals in environments are increased continuously which can cause toward food chain in the environment and become a major human health hazard. Heavy metals are one of the most serious environment pollutants so that it can be derived from both direct sources such as industrial effluents, sludge dumping and indirectly through highway runoffs. As results of these problems, the great interest in metal-microbe interactions have arisen by the researcher and also industrialists to find the suitable methods for solving /or recovering and stabilization of the heavy metals in seawaters, soil and effluents [34].

Conventional methods for removing heavy metals contamination: Numerous clean-up techniques have been suggested and practiced for the removal of heavy metals from the contaminated or pollutant area by using chemical, physical and biological methods. There are several conventional technologies such as precipitation, ion-exchange, electrolytic technologies, chemical extraction, leaching, hydrolysis, polymer micro-encapsulation, and the most common practiced excavation and land filling [35,36]. All of these chemicals methods are posed a serious health and ecological threats due to their toxicity and mutagenicity. Vapour extraction, stabilization, solidification, verifications and membrane technology are previously used for removed the heavy metal ions from pollutant area [36,37].

However, most of these techniques are very expensive for implementation large scale and also dangerous for constant monitoring and control because sometimes it cannot completely remove the heavy metals contaminated. It tend to remove all microbial biota including the useful symbionts such as nitrogen-fixing bacteria as well as other fauna

during the process of sanitisation thereby reducing the biodiversity of the area [38]. These techniques may not always be feasible as their non-specific metals-binding properties [39].

Disadvantages of conventional methods: The high cost of technologies have always used for entirely changing their manufacturing processes or most of the industrialists are not implementing clean-up technologies or replacing their old systems with cleaner, safer and environmental friendly machinery. The use of conventional chemicals for treating heavy metals pollution cannot economically feasible, especially when dealing with low metal-ion concentrations.

According to the Huang et al. [40], several disadvantages in this method can be found. The erratic metal ions removal, high reagent requirements and the generation of toxic sludge are frequently difficult to dewater and need extreme caution in their disposal. Yazdani et al. [41] reported that the conventional methods are expensive especially for handling large amount of water and wastewater contains heavy metals in low concentrations. The needs of several innovative treatment technologies for the removal of heavy metals ions from wastewater are required. Khan et al. [35] suggested that there are possibilities of employing the technology using biological treatments or bioremediation techniques as the alternative methods for removing the heavy metals ions from the contaminated soils or waters.

Bioremediation

Bioremediation is considered alternative processing methods for removing the heavy metals ions from polluted area. Bioremediation is naturally living organisms to reduce the environmental pollutants into less toxic forms. It is followed by bacteria and fungi or plants to degrade or detoxify hazardous ingredients to human health /or the environment. Huang et al. [40] have defined bioremediation as a process by which organic or inorganic waste biologically degraded or transformed usually to innocuous materials. The process can function naturally or can be enhanced by adding an electron acceptor, nutrient or other factors. The microorganisms may be isolated from an indigenous contaminated area or elsewhere and apply to the contaminated site. Contaminant materials are transformed by living organisms through reactions that take place as a part of their metabolic processes. The use of microbial metabolism process has offered a viable, safer, more efficient and less expensive for cleaning of pollutions.

The principles of the bioremediation can be divided into several techniques that including biofilters, bioventing, biosorption, composting, bioaugmentation, bioreactor, land farming and biostimulation [40]. Khan et al. [35] pointed out that the control and optimization of bioremediation processes are complex factors. These factors include the presence of microbial population proficient of degrading the pollutants, the availability of contaminants to the

Heavy Metals	Sources	Effects and Significance
Arsenic	Mining by product, pesticides, chemical waste.	Toxic and possibly carcinogenic.
Beryllium	Coal, nuclear power and space industries.	Acute and chronic toxicity.
Boron	Coal, detergent formulations, industrial waste.	Toxic to some plants.
Chromium	Metal plating, cooling tower water additive.	Essential trace element, possibly carcinogenic as Cr (VI).
Iron	Corroded metal, industrial wastes.	Essential nutrient (component of haemoglobin), damages materials.
Lead	Industrial sources, mining, plumbing, fuels.	Toxicity, wild destruction.
Manganese	Mining, industrial waste, acid mine drainage	Toxic to plants at higher levels.
Mercury	Industrial waste, mining, coal.	Acute and chronic toxicity.
Molybdenum	Industrial waste, natural sources, cooling-tower water additive.	Toxic to animals, essential for plants.
Selenium	Natural geological sources, sulphur, coal.	Essential at low levels, toxic at higher levels, causes 'alkaline disease'.

Table 2: Sources and effects of heavy metals in natural water [22].

microbial population and the environment factors as like as soil type, temperature, pH, the presence of oxygen or other electron acceptors, and nutrients. Bioremediation are unique method for cleaning the polluted environments from the atmosphere (industrials emissions and soil vent gases), solids (soils, sediments and also sludge), liquids (groundwater, industrial effluents) and raw materials from industrial processing. Living or non-living microorganisms can use their enzymes to accomplish in the task [36,42].

Several researchers have reported that some potential microbes are able to tolerate with heavy metals either they able to remove them from the environments or break them down to less toxic or completely benign forms then utilize in their metabolic processes for growth [36,42]. Microbial resistance and tolerance toward pollutants particularly heavy metals are absolutely vital in the bioremediation processes as required microorganisms such as fungi and bacteria that come into closer physical contact with the pollutants due to accomplish the processes and tasks [43,44]. Joho et al. [45] have reported that the heavy metals resistance can be divided into two main categories that are:

The accumulation of metal ions can be reduced by the cells because of the elimination of metals chelating substances or by the breakdown of the specific transport system.

Changes in the intracellular distribution of the ions by binding to specific intracellular molecules.

The processes of bioremediations are involved in metals-microbe interaction including methods of adsorption, complexation, precipitation, oxidation and reductions. The chemical reactions between microorganisms and metal ions are categorized into six distinct processes [26,46], namely (i) intracellular accumulation, (ii) cell wall associated metals, (iii) extracellular mobilization or immobilization of metals, (iv) metalsiderpohore interactions, (v) extracellular polymer-metals interaction with transformation and (vi) volatilization of metals. Kapoor and Viraraghavan [47] indicated that the uptake of heavy metals by the microorganisms could be taken place the toxic ions in several ways in which the bacteria, fungi and algae [17,48]. Overview of bioremediation potential for heavy metals contamination through microbe-heavy metals interactions are shown in Table 3.

Biosorption

Biosorption is an important for researcher to explore microbes from the ecological environment for the use in metal. Lovely [49] is manipulated the microbe-metals interactions including reduction for anaerobic respiration, detoxification, biosorption, bioleaching, bioaccumulation and biomineralization. Biosorption process is extensively studied *via* microbial biomass as a biosorbent for heavy metal removal.

Biosorption is the process that includes the uses of microbes to detoxify and control the environments pollutants. Based on

the interaction between living and non-living microorganisms, and metallic ions in the systems are cleaned up the polluted sites. Accordingly, both living and dead biomass can be entirely employed in biosorptive processes, and it is frequently showed clear tolerance towards metals and other conditions such as low pH [17,47,48]. The combination pathway or genetically modifications of microorganisms have the ability to reduce particular toxic substances.

The use microbial biomass is effectively removed different types of metals ions concentration from aqueous solutions [17,50]. This method is more cost effective and eco-friendly. There are two ways /or mechanisms for microbes in accumulating the heavy metals; biosorption- a metabolism-independent binding of negatively charged free groups to the fungal cell wall, (b) bioaccumulation, an energy-dependent metal influx [50,51]. As a concern of metal accumulation, microorganisms have developed with different mechanisms of metal resistance, including cell membrane metal efflux [52], intracellular chelation by metallothionein proteins and glutathione-derived-peptides called phytochelatins [53,54] as well as metal compartmentalization in vacuoles [4].

Biosorption Techniques

The biosorption techniques are independently up taken the biological metabolic cycle. These biosorption technologies are alternatively used several natural materials of biological origins which include bacteria, yeast, fungi, and algae [55]. Shumate and Strandberg [56] have defined biosorption as a 'non-directed physic-chemical interaction that may occur between metal or radio nuclide species and cellular compounds of biological species. Kapoor et al. [57] have defined biosorption as an interaction between living and non-living microorganisms, and metallic ions in the system. On the other hand, biosorption is involved the uses of microbes to detoxify and control environmental impurities, and received attention to clean up polluted sites as well as comprehensively studied using microbial biomass as a biosorbent for heavy metals removals [17,18].

The processes of biological components are involved in living or non-living organisms. These processes are not only required active cell metabolism. There are several types of chemical groups that may be removed by these processes which included compound of heavy metals that have strong electronegative such as hydroxyl, sulfhydryl ions, carboxyl in anionic groups, as well as phosphate group and nitrogen containing group like amino groups. All these groups are played a vital role in the accumulation and the binding of many toxic metals in the non-specific or specific binding sites on the cell membrane of the microbes at which it will absorb and turn these toxins into their cellular structures [58]. Biosorption techniques are offered several advantages due to require low operating cost and high efficiency for removing low concentration of heavy metals from wastewater. This method is helped the metal ions to recover, stabilize or buried back after their physically removed from the environments [18,36,57].

Heavy Metals	Mechanisms of microbe contaminant interactions	Types of contaminant alteration
Copper	Sorbs to extracellular polymer and biomass.	Immobilized by sorption process.
Nickel	Sorbs to extracellular polymer and biomass.	Immobilized by sorption process.
Zinc	Sorbs to extracellular polymer and biomass.	Immobilized by sorption process.
Mercury	Enzymatically oxidized, reduced or methylated to promote detoxification.	Volatilized or immobilized by sorption, methylation, and precipitation.
Chromium	Enzymatically oxidized or reduced to promote detoxification.	Immobilized by precipitation.
Manganese	Sorbs to extracellular polymers and biomass.	Immobilized by sorption; methylation possible.
Iron	Sorbs to extracellular polymers and biomass.	Immobilized by sorption; methylation possible.

Table 3: Overview of bioremediation potential for heavy metals contamination through microbe-heavy metals interactions [46].

Filamentous Fungi for the Absorption of Heavy Metals and the Future Cleaning-Up Microorganisms

Filamentous fungi are included as a biosorption agent. It is preferred towards other organisms for the bioremediation due to their ability to remove the concentrated heavy metal ions from liquid substrates. Many fungal species have been reported such as *Trichoderma autoviride*, *T. harzianum*, *T. virens* and *Aspergillus niger*, that are used in the process of cleaning polluted areas [1,17,18,59,60].

The tolerance and capability of detoxify metals are involved numerous mechanisms such as valence transformation, extra and intracellular precipitation and active uptake. Among the above reasons, it is considered as potential alternative synthesis resins for the remediation of dilute solution of metals and solid wastes. It has versatile biosorption group; can grow and work under extreme conditions of pH, temperature and nutrient availability as well as high metal concentrations.

The resistances of the fungi toward heavy metals can define as the ability of organisms are survived metals toxicity by means of mechanism formed in direct response to metal species concerned. Compared to other biosorption agent, fungi biomasses have high percentage of cell wall materials that are involved superb metal binding properties in which it can take considerable quantities of heavy metals even the absence of physiological activity.

Interactions of fungi with metals to detoxify heavy metals ions from the polluted sites may occur through several mechanisms (Figure 1) [61-63]:

Mobilization of metals

This process occurs due to the production and excretion of fungal products such as citric acid, an efficient metal ions chelator and oxalic acid which interact with metal ions to produce insoluble oxalate, resulted from the dissolution of primary metals containing phosphate. These organic acids may increase the metal solubility by means of mycosphere acidification and production of metal-complex structure.

Bio-sorption to cell wall

Fungal cell wall is the first cellular components that interact with metal species, thus play an important roles as a protective layer and barrier that control the uptake of potentially toxic metals into the cell. Interaction of metal with fungal cell wall involves a complex mechanism which includes several processes such as ion exchange, complexation, crystallization, adsorption and precipitation and also influenced by the biomass concentration and chemical behaviour of the metals.

Metal uptake

Specific transporters are responsible or the uptake of essential metals. These transporters are possible to react with other type of metals. Carriers may consist of all the metabolically-coupled and H⁺-gradient driven transport system.

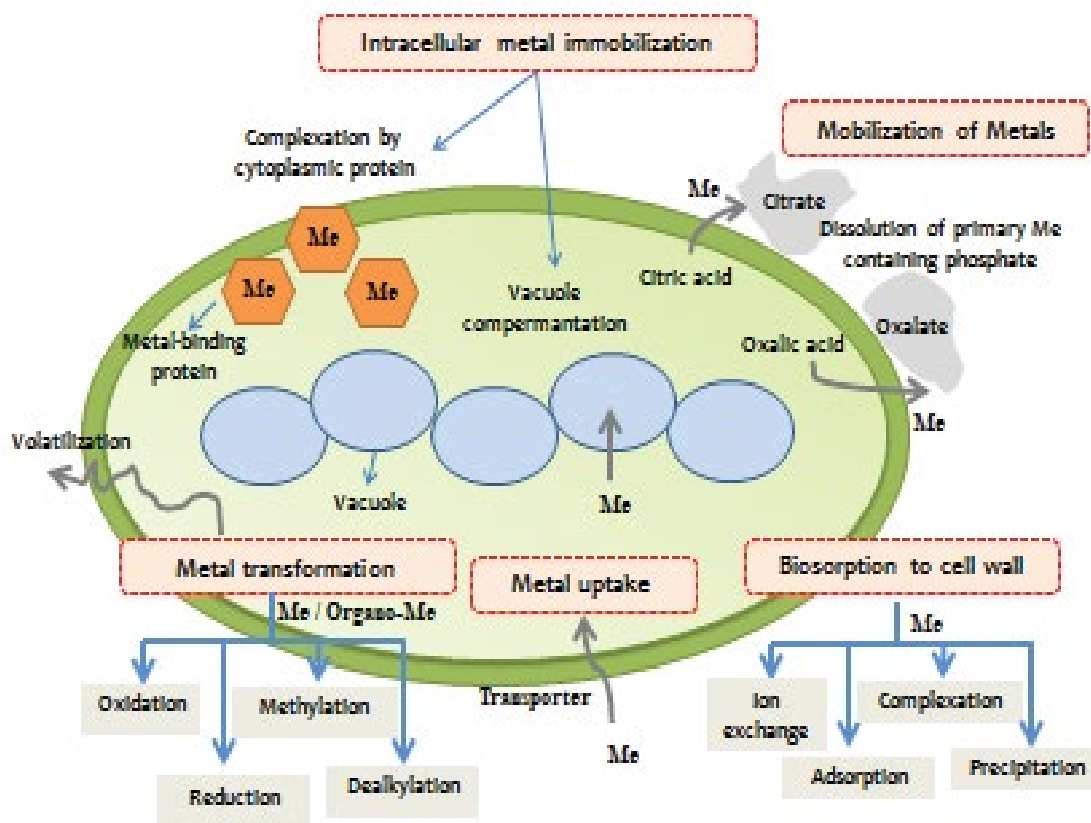


Figure 1: Schematic of fungi-metal interactions (Adapted from [61-63]).

Intracellular metal immobilization

Intracellular metal immobilization involves two processes that is vacuoles compartmentation and complexation by cytoplasmic protein, called metallothioneins and phytochelatins, a rich SH peptide. Fungal vacuole plays important roles in molecular degradation, storage of metabolites, regulation of cytosolic concentrations of metal ions and detoxifies potentially toxic metal ions. Metallothioneins is a metal-binding protein that can modulate the intracellular concentrations and bind both the essential metal such as Cu and Zn and inessential metal such as Cd.

Metal transformations

Fungi can facilitate biotransformation of metals through chemical reactions such as oxidation, reduction, methylation and dealkylation. These reactions may lead to metal volatilization and reduce the metal toxicity. Metals may also transfer to the other parts of the fungi mycelium and plant symbionts by cytoplasmic vesicles and vacuoles.

Kapoor et al. [57] have stated that living and dead cells of fungi are capable to eliminate the heavy metals ions from aqueous solutions. Fungal microorganisms are uptaken the heavy metals ions and offered an alternative method for their removal metals from wastewater. The fungal biomass is easier and cheaper than the removal of heavy metals pollutant by the conventional or traditional adsorbent techniques using activated carbon, coal or ion exchange [36]. Continuously increasing in the used of fungal in numerous biotechnological processes are widely recognized especially in fermentation and bioremediation industries which are included the production of antibiotics, enzymes and industrial acids.

Fungi are used in several industrial fermentation processes because it can serve as an economical and nonstop supplying biomass for removing the metals ions in polluted areas [47]. The ability of the fungi is simply grown in considerable amounts using unsophisticated fermentation techniques and inexpensive growth media. Some researcher are informed that the metal uptake mechanisms of free-living fungi are involved three distinct processes which are included the extracellular uptake through an ion exchange process, intracellular accumulation and the trapping of particulates rich in metals contents [1,41].

Siddiquee et al. [60] found good agents for biosorption of heavy metal ions using isolates of *Trichoderma autroviride*, *T. harzianum*, and *T. virens*. The selected strains of fungi are potential candidate in the research area due to its frequent presence in high polluted area [1,41]. Filipovic et al. [64] used *Aspergillus niger* strain and found better biosorption capabilities with different heavy metals ions of Cu^{2+} , Zn^{2+} , and Ni^{2+} , at pH ranging from 4 to 6. In similar studies done by Michael et al. [39] and reported that *A. niger* is able to grow on culture plates amended with heavy metals and showed five times better inhibition than the growth of yeast. So *Aspergillus niger* biomass has showed the removal of heavy metals in orderly 70% for Zn and 91% for Cu from the wastewater.

Siddiquee et al. [60] mentioned that the resistance levels of different concentrations of heavy metals are diverse because the uses different strains of filamentous fungi of *T. aureoviride*, *T. harzianum*, and *T. virens*. Based on their results, the *T. virens* strain T128 gave the highest tolerance ability for Ni^{3+} and Pb^{2+} in a 1200 mg/L concentration. The accumulation and uptake capacity is determined by the maximum removal of Pb^{2+} , Cu^{2+} , and Ni^{3+} by a *T. harzianum* in liquid medium compared to other fungi. The metal removal occurred of 13.48 g/g for

Pb^{2+} , 3.1254 g/g for Cu^{2+} and 0.8351 g/g for Ni^{3+} , respectively. For Zn^{2+} , the highest tolerance and uptake capacity of metal is recorded at 3.1789 g/g by *T. virens*. The mycelium of *Rhizopus* is an excellent biosorbent towards lead, cadmium, copper, zinc and uranium [4]. *Mucorales* species is described as good biosorbents [65] and *Fusarium flocciferum* is used to remove cadmium and nickel from industrial effluents [66].

Karcprzak and Malina [59] have stated that filamentous fungi have the capacity to form chemical complexes between the metal ions and extracellular enzymes, besides the ability to bind the metals ions to their cell walls. Fungi are accumulated broad range of metals such as Pb, Cd, Cu, Zn, and Ni. Due to the composition of their cell walls that are obtained of polysaccharides, protein and lipids are accumulated substantial amounts of metals, so, they can provide an array of binding sites for metals ions.

Fungal cell walls and their components are played a key role in biosorption process. Kapoor et al. [57] have reported that the high volume of the microorganisms and their ability of detoxify metals among the fungi are considered as the potential alternative synthetic resins for removing the diluted solutions of metals and solid wastes. However, it is not only the cleaning process but also protect the environments and biodiversity as well as allowing for the recovery of the metals and their subsequent reuses [3,36,67].

Fungi have a versatile group, for example they can adapt and grow under varying conditions of pH, temperature and nutrient availability as well as at high metals concentration [36,41,68]. Fungi are one of the most appropriate organisms used to bioremediation. Because, they can tolerate varies environmental and toxic conditions such as a higher concentration of metals levels, and lower pH condition also. They have the ability and capacity to bind with heavy metals to their cell walls which are enhanced the intracellular accumulation of those toxins. Consequently, the response of these microorganisms towards toxic heavy metals concentrations are an important due to their interest in the reclamation of contaminated areas. The ability of selected fungi strains towards remediation of heavy metals ions are evaluated by characterizing the bioaccumulations of these metals.

Tolerance of Selected Heavy Metals by *Trichoderma* sp

Many past researches on the tolerance and biosorption involving characterization and identification of heavy metals resistant fungi have been carried out (Table 4). Screening and identification of metal resistant fungi can be done by fine methodology for tolerance of the fungal isolates towards the heavy metals. It can be measured as minimum inhibitory concentration (MIC) which indicates the lowest concentration of metal that can prevent growth of the isolate. The MIC can be determined based on the biomass dry weight (%). The fungi biomass growth pattern represents the adaptation of the fungi to the heavy metals in which the growth will be reduced as the metal concentration increases [69].

Most of the *Trichoderma* species have high ability of the resistance and tolerate with heavy metals [17,59,60]. As reported by Townsley and Ross [70,71] and Townsley et al. [72], *T. viride* has showed the similar uptake pattern with *Aspergillus niger* and *Penicillium spinulosum*. This fungal is able to remove varying heavy metals from aqueous solution through ion-exchange reaction uptake. Studies done by Townsley et al. [73] and Ross and Townsley [74] found that copper, gold, zinc, cadmium and manganese are able to remove the heavy metals ion by fungi. They are found very impressive results with an average of 90% of heavy metals uptake achieved within 10 minutes by fungal cell. The

Fungi	Heavy Metals Resistant	Area	References
<i>Trichoderma atroviride</i>	Copper, Zinc, Cadmium	Sewage Sludge, Madrid, Spain	[1]
<i>Trichoderma atroviride</i> , <i>Mortierella exiguua</i>	Zinc, Barium, Iron	Tarnowskie Gory, South Poland	[59]
<i>Aspergillus niger</i> , <i>Penicillium</i> , <i>Alternaria</i> , <i>Rhizopus</i> , <i>Monilia</i> , <i>Trichoderma</i>	<i>Aspergillus</i> , <i>Penicillium</i> , <i>Geotrichum</i> , <i>Alternaria</i> , <i>Fusarium</i> , <i>Rhizopus</i> , <i>Monilia</i> , <i>Trichoderma</i>	Aligarh, Uttar Pradesh, India	[17]
<i>Aspergillus niger</i> , <i>Trichoderma asperellum</i> , <i>Penicillium simplicissimum</i>	Copper, Lead	Langat River, Selangor, Malaysia	[18]
<i>Trichoderma atroviride</i>	Zinc	Serdang Industrial Area, Selangor	[41]
<i>Trichoderma viride</i> , <i>Aspergillus niger</i> , <i>Penicillium spinulosum</i>			[66-68]
<i>Trichoderma viride</i> , <i>Aspergillus oryzae</i> , <i>Aspergillus niger</i> , <i>Mucor racemosus</i> , <i>Penicillium chrysogenum</i>			[72,73]
<i>Trichoderma</i> , <i>Aspergillus</i> , <i>Mortierella</i> , <i>Apecilomyces</i> , <i>Penicillium</i> , <i>Pythium</i> and <i>Rhizopus</i>	Cobalt	Serpentine soil of Andaman	[74]
<i>Trichoderma atroviride</i>	Copper, Zinc, Cadmium	Madrid	[87]
<i>Trichoderma</i> (FA-06)	Arsenic	West Bengal, India	[79]
<i>Trichoderma atroviride</i> strains F6	Cadmium, Nickel	Unspecified	[80]
<i>T. harzianum</i> , <i>F. phyllophilum</i>	Zinc, Plumbum	Sicily, Italy	[81]
<i>T. virens</i> (PDR-28)	Arsenic, Copper, Cadmium, Nickel, Plumbum, Zinc	Mine tailing soil, South Korea	[82]

Table 4: *Trichoderma* sp. their metal of tolerance based on past researches.

metal accumulations of growing cells are varied within the cell age. Maximum metal uptake during the lag phase or known as early stages of growth and declined as cultures reached a stationary phase.

During the biosorption processes are changed in pH recorded. pH is dropped during the growth and the reduction in metal-uptake pattern. In addition, the changes in cell wall composition with growth and the release of metabolites that are binder with the metal ions during the constant pH recorded [73,74]. Huang et al. [75] have recorded the biosorption of cadmium (Cd) on various fungal strains are pH very sensitive. Some fungal strains have a better biosorption capacity in the acidic range.

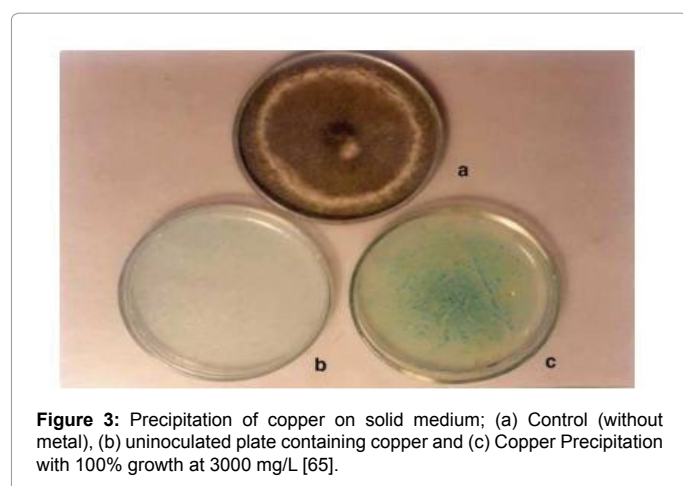
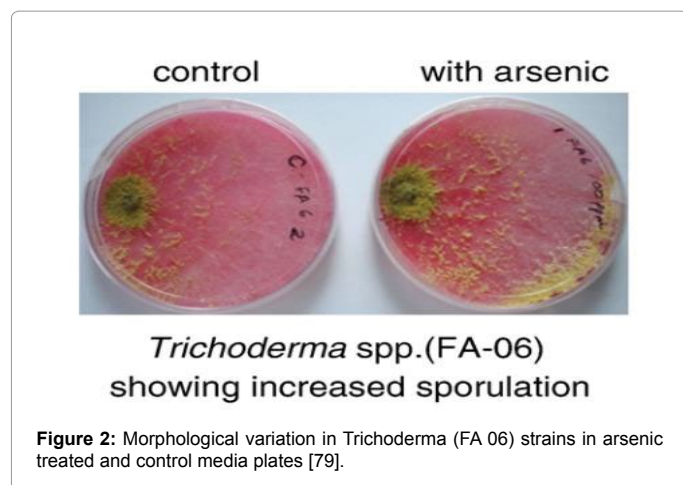
Trichoderma viride has showed higher amount cadmium uptaken at lower concentration of biomass comparing to *A. oryzae*, *A. niger*, *Mucor racemosus* and *P. chrysogenum* [76,77]. The higher biomass concentrations can be attributed to the electrostatics interactions of the functional groups at the cell surfaces. Higher concentrations in cell suspensions are attached each other and thus lower the cell surface area in contact with the solution. The growth conditions can affect the metal uptake of the biomass. *Trichoderma* fungi have diverse metabolic adaptability and showed several heavy metals resistance [78]. Several filamentous fungi (*Aspergillus*, *Mortierella*, *Apecilomyces*, *Penicillium*, *Pythium* and *Rhizopus*) are isolated from serpentine soil of Andaman for cobalt resistance. All of these fungi from metal-percoated environment are similarly efficient in biosorbing Co (II) ions.

Lopez and Vazquez [1] have reported that *T. atroviride* strain isolated from sewage sludge water treatment plant in Madrid. It has ability to survive in high metal concentrations as the natural selection of resistant cells. *In vitro* assays, *T. atroviride* is confirmed a high tolerance against copper, zinc and cadmium. Microorganisms isolated from natural contaminated with heavy metals environments are found high tolerance adaptation of multiple environmental conditions [79]. The responses of micro-organisms may differ from their response to individual metals or/ multiple metals and additive, synergistic or antagonist interactions may occur between metals [80]. Combinations of several ions are affected based on the metal uptake. The selective accumulations of heavy metals by microorganisms are determined by interionic competition in which metal cations compete for the binding sites on the cell wall [69,81,82]. Metal uptake has assayed through by the fungus is cultured in the existing of a single metal and in the

presence of a combination of two or three cations, where additive and synergistic interactions are observed. Combinations of zinc, copper and cadmium are assayed as single metals (50 mg/ml) moderately inhibited the growth of *T. atroviride*.

Trichoderma has found among the other fungi with highest arsenic effectively removal from arsenic contaminated agricultural soils [83]. Arsenic is a toxic compound which existed in inorganic and organic forms. Different strains have different ability in bioindicators of arsenic pollution. *Trichoderma* strain FA-06 is able to sporulate well on the arsenate exposure as compared to control and other *Trichoderma* strains [84]. Some *Trichoderma* strains have showed a decrease in pH ranging from 12.03 to 18.05% and 24.08 to 34.24% in arsenic treatment (Figure 2 and 3). They have concluded that *Trichoderma* is the best capability in biovolatilizing arsenic and efficient in removing arsenic from liquid medium by 56.12%. *T. atroviride* strain F6 significantly alleviated the cellular toxicity of cadmium and nickel to plants [84]. Inoculations of *B. juncea* (L.) Coss. var. *foliosa* with *T. atroviride* strain F6 have showed the increase in fresh weight in Cd for 110%, Ni for 40% and Cd-Ni for 170%, contaminated soils, respectively (P<0.05). The translocation factors and metal bioconcentration factors are calculated for the inoculated plant increased comparing to the non-inoculated plants. The results are indicated that the efficiency of phytoextraction for *B. juncea* (L.) Coss. var. *foliosa* has enhanced after inoculating with *T. atroviride* strain F6. The fungal treated plants grown in Cd-Ni combination contaminated soils are showed higher phytoextraction efficiency than those in Cd or Ni contaminated soils. So, it is suggested that the fungus *T. atroviride* strain F6 endowed with organic-degrading capabilities could be exploited for fungi-assisted phytoremediation of mixed organic-metal contaminated soils.

Four different types of fungal have ability on growing in the presence of heavy metals and monitored their cysteine and glutathione content and the activity of O-acetylserine (thiol) lyase (OASTL) which is involved in cysteine biosynthesis studies [85]. They have found that the presences of Zn and Pb have not affect the fungal growth or sporulation at the concentration used of 5 and 10 ppm. Cysteine and glutathione contents are always higher when fungi are grown in the presence of toxic metals. As *T. harzianum* and *F. phyllophilum* are showed the best growth rate on Cd and Hg. *T. harzianum* and *F. pylllophilum* have grown in the presence of Zn, Pb, Cd, and Hg and accumulated high amounts of these metals when metal-containing media.



Babu et al. [86] have successfully isolated *T. virens* named PDR-28 and found MICs for 1300 mg of As, 1000 mg of Cu, 1300 mg of Cd, 700 mg of Ni, 2500 mg of Pb and 3100 mg of Zn have indicated the heavy metal adaptation stress; differences in tolerance may reflect different adaptation strategies or mechanisms involving permeability barriers, intra and extracellular sequestration, efflux pumps, enzymatic detoxification, and metal speciation. Anand et al. [68] and Iskandar et al. [18] have identified *T. asperellum* and *T. viride* at heavy metal contaminated sites. Strain PDR-28 effectively removed heavy metals in the order of Pb>Cd>As>Zn>Cu from liquid media containing 100 mg heavy metals L⁻¹. Strain PDR-28 may be more effective heavy metal biosorbent at contaminated sites and in industrial wastewater. Several strains of *Trichoderma* such as *T. atroviride*, *T. harizianum*, and *T. viride* have been used for removing heavy metals from aqueous solution [18,60,68].

Trichoderma viride is tolerated of 500 mg Cu L⁻¹ in modified Czapek Dox minimal agar medium reported by Anand et al. [68]. In agar medium, effect of 1000–5000 mg/L of Cu(II) are tested and the results found of 100% growth of *T. viride* upto the concentration of 3000 mg/L against control (Figure 2). Similar level of tolerance reported by Tsekova and Todorova [87] by using *A. niger* B-77 strain where 300 mg/L of Cu(II) ion is inhibitory to the growth of the organism. The mycelia turned blue both on agar media and in broth culture at all concentrations of copper, which is the binding of Cu(II) to the fungal cell wall. Blue coloured mycelia reported by Subramanyam and

Gupta [88] and Venkateswerlu et al. [89] in the presence of copper in *Neurospora crassa* and *Cunninghamella blackesleeana*, respectively. They suggested that the blue color of the mycelia is binded of Cu(II) with cell wall protein of the mycelium. Interestingly, 81.78% of the Cu(II) removal is observed at a copper concentration of 100 mg/L in 72 h. Gadd and Griffiths [90,91] and Townsley and Ross [70] have reported the maximal copper uptake in lag phase in *Aureobasidium pullulans* and *A. niger*, respectively.

Iskandar et al. [18] have used four strains of fungus, namely, *A. niger*, *A. fumigatus*, *T. asperellum*, and *Penicillium simplicissimum*. *T. asperellum* has able to tolerate with 800 mg Cu L⁻¹ and 1000 mg Pb L⁻¹ in PDA medium, showing a better uptake capacity for Pb. This result indicated that *T. asperellum* is a promising biosorptive agent. Among the filamentous fungi of *Aspergillus*, *Penicillium*, *Geotrichum* *Alternaria*, *Fusarium*, *Rhizopus*, *Monilia* and *Trichoderma* are identified in soil previously reported by Zafar et al. [17]. All strains are tested for the Cobalt, chromium, copper, cadmium and nickel. *Trichoderma* is showed MIC of heavy metals of the average of 6 to 3 mg ml⁻¹.

The exposure of fungi on zinc resistance with different concentrations between 0 – 6000mg/L has conducted by Yazdani et al. [41]. Among 30 isolates fungi cultured, only *T. atroviride* is found able to grow at 6000 mg/L of zinc concentration on PDA. Accordingly, *T. atroviride* is the highest tolerance towards Zn. Uptake capacity of *T. atroviride* is ranged from 18.1–26.7 mg/g in liquid media at Zn concentrations from 500–1000 mg/L. *T. atroviride* showed of 47.6–64% adsorption in liquid media and 30.4–45.1% of absorption for Zn. Based on this study, 5.7–7.4% of Zn removal is observed due to biomass washing. The high adsorption, relatively low absorption and high uptake capacity of Zn suggested that *T. atroviride* is a potential bioremediator of Zn. This result has proven that the filamentous fungi isolated from soil contaminated heavy metals sites have more potential strains comparing to other located isolates fungi and it can be exploited for removing heavy metal ions from aqueous solution.

Conclusion

Interest in processes involving heavy metal uptake by micro-organisms has increased considerably in recent years due to the biotechnological potential of micro-organisms in removing and/or recovery of metals. The conventional methods such as synthetic ion exchangers are considered as mature technologies. Biosorption is still in its developmental stages, and additional improvements in both performance and costs can be expected. Based on the previous research on the capability of *Trichoderma* sp. in removing heavy metals, it is completely suggested that *Trichoderma* sp. should be considered as the agents in bioremediation process.

References

1. López Errasquín E, Vázquez C (2003) Tolerance and uptake of heavy metals by *Trichoderma atroviride* isolated from sludge. *Chemosphere* 50: 137-143.
2. Dedyukhina EG, Eroshin VK (1991) Essential metal ions in the control of microbial metabolism. *Proc Biochem* 26: 31-37.
3. Gadd GM (1986) Fungal responses towards heavy metals. In: Herbert RA, Codd GA, *Microbes in Extreme Environments*. Academic Press, London.
4. Volesky B (1994) Advances in biosorption of metals: selection of biomass types. *FEMS Microbiol Rev* 14: 291-302.
5. Domenech X (1998) *Química Ambiental El Impacto Ambiental de los Residuos*. Miraguano Ediciones, Madrid.
6. Price MS, Classen JJ, Payne GA (2001) *Aspergillus niger* absorbs copper and zinc from swine wastewater. *Bioresour Technol* 77: 41-49.

7. Cochrane WW (1958) *Farm Prices: Myth and Reality*. St Paul. University of Minnesota Press.
8. Williams J, Clarkson JM, Mills PR, Cooper RM (2003) A selective medium for quantitative reisolation of *Trichoderma harzianum* from *Agaricus bisporus* compost. *Appl Environ Microbiol* 69: 4190-4191.
9. Harman GE, Howell CR, Viterbo A, Chet I, Lorito M (2004) *Trichoderma* species—opportunistic, avirulent plant symbionts. *Nat Rev Microbiol* 2: 43-56.
10. Benítez T, Rincón AM, Limón MC, Codón AC (2004) Biocontrol mechanisms of *Trichoderma* strains. *Int Microbiol* 7: 249-260.
11. Kredics L, Antal Z, Manczinger L, Nagy E (2001) Breeding of mycoparasitic *Trichoderma* strains for heavy metal resistance. *Lett Appl Microbiol* 33: 112-116.
12. Kredics L, Doczi I, Antal Z, Manczinger L (2001) Isolation and characterization of heavy metal resistant mutants from mycoparasitic *Trichoderma* strains. *Biol. Cont. Fungal Bacterial Plant Pathogens. IOBC wprs Bulletin* 24: 233-236.
13. Radojevic M, Abdullah MH, Aris AZ (2007) *Analisis Air*. Scholar Press, Selangor.
14. Yu MH (2001) *Environment Toxicology: Impacts of Environmental Toxicants on Living Systems*. (3rd Edn) CRS Press LLC, Florida.
15. Forstner U, Wittmann GTW (1983) *Metal Pollution in the Aquatic Environment* (2nd Edn), Springer-Verlag, Berlin.
16. Poli A, Salerno A, Laezza G, di Donato P, Dumontet S, et al. (2009) Heavy metal resistance of some thermophiles: potential use of alpha-amylase from *Anoxybacillus amylolyticus* as a microbial enzymatic bioassay. *Res Microbiol* 160: 99-106.
17. Zafar S, Aqil F, Ahmad I (2007) Metal tolerance and biosorption potential of filamentous fungi isolated from metal contaminated agricultural soil. *Bioresour Technol* 98: 2557-2561.
18. Iskandar NL, Zainudin NA, Tan SG (2011) Tolerance and biosorption of copper (Cu) and lead (Pb) by filamentous fungi isolated from a freshwater ecosystem. *J Environ Sci (China)* 23: 824-830.
19. Arora M, Kiran B, Rani S, Rani A, Kaur B, et al. (2008) Heavy Metals accumulation in vegetables irrigated with water from different sources. *Food Chem* 111: 811-815.
20. Hardman DJ, McEldowney S, Waite S (1993) *Pollution: Ecology and Biotreatment*. Longman Scientific and Technical, England.
21. Mengel K, Kirkby EA (1979) *Principle of Plant Nutrition* (2nd Ed) International Potash Institute Berne, Switzerland.
22. Manahan SE (1997) *Environmental Science and Technology*, Lewis Publishers, New York.
23. Radhika V, Subramanian S, Natarajan KA (2006) Bioremediation of zinc using *Desulfotomaculum nigrificans*: bioprecipitation and characterization studies. *Water Res* 40: 3628-3636.
24. Fosmire GJ (1990) Zinc toxicity. *Am J Clin Nutr* 51: 225-227.
25. Chang R (2005) *Chemistry*, (4th Edn) McGraw-Hill, New York.
26. Davies DJA, Bennett BG (1983) *Exposure Commitment Assessment of Environment pollutants Vol. 3* MARC (Monitoring and Assessment Research Centre) Report Number 30 MARC Publication, London.
27. Batey T, Berryman C, Line C (1972) The Disposal of Copper-Enriched Pig Manure Slurry on Grassland. *J Br Grassland Soc* 27: 139-143.
28. Lee KS (2003) *Kajian Kepekatan Logam Berat Kuprum dan Zink dalam sedimen Sungai Kimanis, Sabah*.
29. Handy RD (2003) Chronic effects of copper exposure versus endocrine toxicity: two sides of the same toxicological process? *Comp Biochem Physiol A Mol Integr Physiol* 135: 25-38.
30. Insel P, Elainturner R, Ross D (2004) *Nutrition*, (2nd Edn) American Dietetic Association, New York.
31. Dunnick JK, Elwell MR, Radovsky AE, Benson JM, Hanh FF, et al (1995) Comparative Carcinogenic Effects of Nickel Subsulfide, Nickel Oxide or Nickel Sulphate Hexahydrate Chronic Exposure in the Lung. *Cancer Res* 55: 5251-5256.
32. Crosby DG (1998) *Environmental Toxicology and Chemistry* Oxford University Press, New York.
33. Blackmore R, Reddish A (1996) *Global Environmental Issues* Hodder and Stoughton, United Kingdom.
34. Sani RK, Peyton BM, Brown LT (2001) Copper-induced inhibition of growth of *Desulfovibrio desulfuricans* G20: assessment of its toxicity and correlation with those of zinc and lead. *Appl Environ Microbiol* 67: 4765-4772.
35. Khan AG, Bari A, Chaudhry TM, Qazilbash AA (1997) Phytoremediation-Strategy to Decontaminate Heavy Metal Polluted Soils and to Conserve the Biodiversity of Pakistan Soils. In: Mufti SA, Woods CA, Hasan SA, Biodiversity of Pakistan Pakistan Museum of Natural History, Islamabad and Florida Museum of Natural History, Gainesville, Florida.
36. Qazilbash AA (2004) Isolation and Characterization of Heavy Metal Tolerant Biota from Industrially Polluted Soils and Their Role in Bioremediation. *Biological Sci* 41: 210-256.
37. Balba MT, Al-Awadhi N, Al-Daher R (1998) Bioremediation of oil-contaminated soil: microbiological methods for feasibility assessment and field evaluation. *J Microbiol Meth* 3: 155-164.
38. Chaudhry TM, Hill L, Khan AG (1999) Bioremediation of Iron and Zinc contaminated dumped filter cake by microbes, plants and associated mycorrhizae. In: Wong M, Bakar AJM, Bioremediation of heavy metals, CRC Press, Boca Raton, Florida.
39. Price MS, Classen JJ, Payne GA (2001) *Aspergillus niger* absorbs copper and zinc from swine wastewater. *Bioresour Technol* 77: 41-49.
40. Huang JP, Huang CP, Morehart AL (1991) Removal of heavy metals by fungal (*Aspergillus oryzae*) adsorption. In: Vernet JP, Heavy Metals in the Environment, Elsevier, London.
41. Yazdani M, Chee KY, Faridah A, Soon GT (2010) An in vitro study on the Adsorption, Absorption and uptake Capacity of Zn by the Bioremediator *Trichoderma atroviride*. *Environ Asia* 3: 53-59.
42. Atlas RM, Unterman R (1999) *Bioremediation in Manual of Industrial Microbiology and Biotechnology*, 2nd American Society for Microbiology Press, Washington DC.
43. Parales RE, Ditty JL, Harwood CS (2000) Toluene-degrading bacteria are chemotactic towards the environmental pollutants benzene, toluene, and trichloroethylene. *Appl Environ Microbiol* 66: 4098-4104.
44. Sharma PK, Balkwill DL, Frenkel A, Vairavamurthy MA (2000) A new *Klebsiella planticola* strain (Cd-1) grows anaerobically at high cadmium concentrations and precipitates cadmium sulphide. *Appl Environ Microb* 66: 3083-3087.
45. Joho M, Inouhe M, Tohoyama H, Murayama T (1995) Nickel resistance mechanisms in yeasts and other fungi. *J Ind Microbiol* 14: 164-168.
46. Ford T, Mitchell M (1992) *Microbial Transport of Toxic Metals: Environmental Microbiology*. John Wiley and Sons, New York.
47. Kapoor A, Viraraghavan T (1998) Biosorption of heavy-metal on *Aspergillus niger*: effect of pretreatment. *Biores Technol* 63: 109-113.
48. Gadd GM (1990) *Fungi and yeast metal accumulation: Microbial Mineral Recovery*, McGraw-Hill, New York.
49. Lovley DR, Phillips EJ (1994) Reduction of Chromate by *Desulfovibrio vulgaris* and its c(3) Cytochrome. *Appl Environ Microbiol* 60: 726-728.
50. Gomes NCM, Mendonca-Hagler L, Savvaids I (1998) Metal bioremediation by microorganisms. *Rev Microbiol* 29: 85-92.
51. Cervantes C, Gutierrez-Corona F (1994) Copper resistance mechanisms in bacteria and fungi. *FEMS Microbiol Rev* 14: 121-137.
52. Kamizono A, Nishizawa M, Teranishi Y, Murata K, Kimura A (1989) Identification of a gene conferring resistance to zinc and cadmium ions in the yeast *Saccharomyces cerevisiae*. *Mol Gen Genet* 219: 161-167.
53. Kneer R, Kutchan TM, Hochberger A, Zenk MH (1992) *Saccharomyces cerevisiae* and *Neurospora crassa* contain heavy metal sequestering phytochelatin. *Arch Microbiol* 157: 305-310.
54. Presta A, Stillman MJ (1997) Incorporation of copper into the yeast *Saccharomyces cerevisiae*. Identification of Cu(I)-metallothionein in intact yeast cells. *J Inorg Biochem* 66: 231-240.
55. Bayramoğlu G, Bektaş S, Arica MY (2003) Biosorption of heavy metal ions on immobilized white-rot fungus *Trametes versicolor*. *J Hazard Mater* 101: 285-300.
56. Shumate ES, Strandberg WG (1985) Accumulation of metals by microbial cells. *Comprehensive Biotechnol* 13: 235-247.

57. Kapoor A, Viraraghavan T, Cullimore DR (1999) Removal of heavy metals using fungus *Aspergillus niger*. *Biores Technol* 70: 95-104.
58. Tobin JM, Cooper DG, Neufeld RJ (1984) Uptake of Metal Ions by *Rhizopus arrhizus* Biomass. *Appl Environ Microbiol* 47: 821-824.
59. Karcprzak M, Malina G (2005) The tolerance and Zn²⁺, Ba²⁺ and Fe²⁺ accumulation by *Trichoderma atroviride* and *Mortierella exigua* isolated from contaminated soil. *Can J Soil Sci* 85: 283-290.
60. Siddiquee S, Aishah SN, Azad SA, Shafawati SN, Naher L (2013) Tolerance and Biosorption capacity of Zn²⁺, Pb²⁺, Ni³⁺ and Cu²⁺ by filamentous fungi (*Trichoderma harzianum*, *T. atroviride* and *T. virens*). *Adv Biosci Biotechnol* 4: 570-583.
61. Harms H, Schlosser D, Wick LY (2011) Untapped potential: exploiting fungi in bioremediation of hazardous chemicals. *Nat Rev Microbiol* 9: 177-192.
62. Valls M, de Lorenzo V (2002) Exploiting the genetic and biochemical capacities of bacteria for the remediation of heavy metal pollution. *FEMS Microbiol Rev* 26: 327-338.
63. Gadd GM (1993) Interactions of fungi with toxic metals. *New Phytol* 124: 25-60.
64. Filipovic KZ, Sipos L, Briski F (2000) Biosorption of chromium, copper, nickel and zinc ions onto fungal pellets of *Aspergillus niger* from aqueous solutions. *Food Technol Biotechnol* 38: 211-216.
65. Remacle J (1990) The cell wall and heavy metals. In: Volesky B, *Biosorption of Heavy Metals*, CRC Press, Boca Raton, Florida.
66. Delgado A, Anselmo AM, Novais JM (1998) Heavy metal biosorption by dried powdered mycelium of *Fusarium flocciferum*. *Water Environ Res* 70: 370-375.
67. Brierley CL, Brierley JA (1993) Immobilization of Biomass for Industrial Application of Biosorption. In: Torma AE, Apel ML, Brierley CL, Warrendale PA, *Biohydrometallurgy Technologies, the Minerals, Metals and Materials Society*.
68. Anand P, Isar J, Saran S, Saxena RK (2006) Bioaccumulation of copper by *Trichoderma viride*. *Bioresour Technol* 97: 1018-1025.
69. Lairini K, Ezzouhri L, Castro E, Moya, and Espinola F (2009) Heavy metal tolerance of filamentous fungi isolated from polluted sites in Tangier, Morocco. *African J of Microbiol Res* 3: 35-48.
70. Townsley CC, Ross IS (1985) Copper uptake by *Penicillium spinulosum*. *Microbios* 44: 125-132.
71. Townsley CC, Ross IS (1986a) Copper uptake in *Aspergillus niger* during batch growth and in non-growing mycelia suspensions. *Experimental Mycol* 10: 281-288.
72. Townsley CC, Ross IS, Atkins AS (1986b) Copper removal from a simulated leach effluent using the filamentous fungus *Trichoderma viride*. In: Eccles HH, Hunt S, *Immobilization of Ions by Biosorption*. Ellis Horwood, Chichester, UK.
73. Townsley CC, Ross IS, Atkins AS (1986) Biorecovery of metallic residues from various industrial effluents using filamentous fungi. In: Lawrence RW, Branion RMR, Ebner HG, *Fundamental and Applied Biohydrometallurgy*. Elsevier, Amsterdam.
74. Ross IC, Townsley CC (1986) The uptake of heavy metals by filamentous fungi. In: Eccles HH, Hunt S, *Immobilization of Ions by Biosorption* Ellis Horwood, Chichester, West Sussex England.
75. Huang CP, Westman D, Quirk K, Huang JP (1988) The removal of cadmium (II) from dilute aqueous solutions by fungal adsorbent. *Water Sci Technol* 20: 369-376.
76. Kiff RJ, Little DR (1986) Biosorption of heavy metals by immobilized fungal biomass. In: Eccles HH, Hunt S, *Immobilization of Ions by Biosorption*. Ellis Horwood, Chichester, UK.
77. Kurek E, Czaban J, Bollag JM (1982) Sorption of cadmium by microorganisms in competition with other soil constituents. *Appl Environ Microbiol* 43: 1011-1015.
78. Pal A, Ghosh S, Paul AK (2006) Biosorption of cobalt by fungi from serpentine soil of Andaman. *Bioresour Technol* 97: 1253-1258.
79. Ashida J (1965) Adaptation of fungi to metal toxicants. *Ann Rev Phytopathol* 3: 153-174.
80. Babich H, Stotzky G (1983) Synergism between nickel and copper in their toxicity to microbes: mediation by pH. *Ecotoxicol Environ Saf* 7: 576-587.
81. Castro F, Viedma P, Cotorás D (1992) [Biomass of *Rhizopus oligosporus* as an adsorbent for metal ions]. *Microbiologia* 8: 94-105.
82. Chang JS, Huang JC (1998) Selective adsorption/recovery of Pb, Cu, and Cd with multiple fixed beds containing immobilized bacterial biomass. *Biotechnol Prog* 14: 735-741.
83. Srivastava PK, Vaish A, Dwivedi S, Chakrabarty D, Singh N, et al. (2011) Biological removal of arsenic pollution by soil fungi. *Sci Total Environ* 409: 2430-2442.
84. Cao L, Jiang M, Zeng Z, Du A, Tan H, et al. (2008) *Trichoderma atroviride* F6 improves phytoextraction efficiency of mustard (*Brassica juncea* (L.) Coss. var. foliosa Bailey) in Cd, Ni contaminated soils. *Chemosphere* 71: 1769-1773.
85. Raspanti E, Cacciola SO, Gotor C, Romero LC, García I (2009) Implications of cysteine metabolism in the heavy metal response in *Trichoderma harzianum* and in three *Fusarium* species. *Chemosphere* 76: 48-54.
86. Babu AG, Shim J, Bang KS, Shea PJ, Oh BT5 (2014) *Trichoderma virens* PDR-28: a heavy metal-tolerant and plant growth-promoting fungus for remediation and bioenergy crop production on mine tailing soil. *J Environ Manage* 132: 129-134.
87. Tsekova K, Todorova D (2002) Copper (II) accumulation and superoxide dismutase activity during growth of *Aspergillus niger* B-77. *Z Naturforsch C* 57: 319-322.
88. Subramanyam C, Gupta PD (1986) Glycogen deposition in *Neurospora crassa* under conditions of copper toxicity: a correlative ultra-structural and biochemical study. *Microbiologica* 45: 55-62.
89. Venkateswerlu G, Stotzky G (1986) Copper and cobalt alter the cell wall composition of *Cunninghamella blakesleeana*. *Can J Microbiol* 32: 654-662.
90. Gadd GM, Griffiths AJ (1977) Microorganisms and heavy metal toxicity. *Microb Ecol* 4: 303-317.
91. López Errasquín E, Vázquez C (2003) Tolerance and uptake of heavy metals by *Trichoderma atroviride* isolated from sludge. *Chemosphere* 50: 137-143.